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CheKine™ Micro Soil Acidic Xylanase (S-ACX) Protease Activity Assay Kit

Cat #: KTB4056 Size: 48 T/48 S 96 T/96 S

[-]	Micro Soil Acidic Xylanase (S-ACX) Protease Activity Assay Kit		
REF	Cat #: KTB4056	LOT	Lot #: Refer to product label
	Detection range: 0.4-2 µmol/mL		Sensitivity: 0.4 µmol/mL
	Applicable sample: Soil		
Å	Storage: Stored at 4°C for 6 months, protected from light		

Assay Principle

Xylanase (EC 3.2.1.8) is mainly produced by microorganisms and can catalyze the hydrolysis of xylan. It is also referred to as pentosanase or hemicellulase, capable of breaking down cell walls in raw materials used in brewing or feed industries, as well as β-glucans. This reduces the viscosity of materials during brewing, promotes the release of effective substances, and decreases non-starch polysaccharides in feed, thereby enhancing the absorption and utilization of nutrients. Due to these properties, xylanase is widely applied in brewing and feed industries. Acidic xylanase (ACX) is generally isolated from acid-tolerant fungi, bacteria, and some molds. CheKine™ Micro Soil Acidic Xylanase (S-ACX) Protease Activity Assay Kit provides a simple, convenient, and rapid method for detecting neutral xylanase activity in soil samples. Its principle is based on ACX degrading xylan into reducing oligosaccharides and monosaccharides under acidic conditions. These products further react with 3,5-dinitrosalicylic acid (DNS) in a boiling water bath to produce a colorimetric reaction with a characteristic absorption peak at 540 nm. The intensity of the reaction solution's color is proportional to the amount of reducing sugars produced during enzymatic hydrolysis. By measuring the increase rate of absorbance at 540 nm, the activity of ACX can be calculated.

Materials Supplied and Storage Conditions

Vit components	Si	Storage conditions	
Kit components	48 T	96 T	Storage conditions
Reagent	18 mL	36 mL	4°C
Reagent II	3 mL	6 mL	4°C, protected from light
Reagent III	13 mL	26 mL	4°C, protected from light
Standard	Powder×1 vial	Powder×1 vial	4°C, protected from light

Note: Before formal testing, it is recommended to select 2-3 samples with large expected differences for pre-experiment.

Materials Required but Not Supplied

• Microplate reader or visible spectrophotometer capable of measuring absorbance at 540 nm



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- 96-well microplate or microglass cuvette, precision pipettes, disposable pipette tips, 1.5 mL EP tube
- · Water bath, centrifuge, 30-50 mesh sieve
- · Deionized water

Reagent Preparation

Reagent I: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C.

Reagent II: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C, protected from light.

Reagent III: Ready to use as supplied. Equilibrate to room temperature before use. Store at 4°C, protected from light.

Note: Reagent III has certain irritation, so personal protection is recommended during use.

Standard: Prepared before use. Add 1 mL deionized water to a bottle, dissolve thoroughly, that is 100 µmol/mL D-Xylose Standard. The remaining reagent can also be stored at 4°C and protected from light for 1 month.

10 μ mol/mL D-Xylose Standard: Take 120 μ L of the 100 μ mol/mL D-xylose standard and add 1,080 μ L of deionized water to prepare a 10 μ mol/mL D-xylose standard solution. Using the 10 μ mol/mL D-xylose standard solution, further dilute the standard according to the table below:

Num.	Standard Volume (µL)	Deionized Water (μL)	Concentration (µmol/mL)
Std.1	200 μL of 10 μmol/mL Standard	800	2
Std.2	150 μL of 10 μmol/mL Standard	850	1.5
Std.3	120 μL of 10 μmol/mL Standard	880	1.2
Std.4	100 μL of 10 μmol/mL Standard	900	1
Std.5	80 μL of 10 μmol/mL Standard	920	0.8
Std.6	40 μL of 10 μmol/mL Standard	960	0.4

Note: Always prepare fresh standards per use; Diluted Standard Solution is unstable and must be used within 4 h.

Sample Preparation

Note: Note: It is recommended to use fresh soil samples.

Fresh soil samples naturally air dried or air dried in an oven at 37°C and sieved through 30-50 mesh sieve.

Assay Procedure

- 1. Preheat the microplate reader or visible spectrophotometer for more than 30 min, and adjust the wavelength to 540 nm, visible spectrophotometer was returned to zero with deionized water.
- 2. Operation table (The following operations are operated in the 1.5 mL EP tube):

Reagent	Test Tube	Control Tube	Standard Tube	Blank Tube
Sample (g)	0.02	0.02	0	0
Reagent (µL)	100	150	0	0
Reagent II (µL)	50	0	0	0

Mix well, incubating at 50°C for 2 h, immediately inactivating at 90°C for 10 min. Centrifuge at 8,000 g for 10 min at room temperature, take the supernatant, and following operations are operated in the new 1.5 mL EP tube:

Supernatant (μL)	100	100	0	0
Standard (µL)	0	0	100	0



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Deionized Water (µL)	0	0	0	100
Reagent III (µL)	100	100	100	100

Mix well, incubate at 90 °C for 5 min, cooling to room temperature with running water, take 180 μ L into 96-well microplate or microglass cuvette, record the absorbance value at 540 nm. The Blank Well is recorded as A_{Blank} , the Standard Well is marked as $A_{Standard}$, the Control Well is marked as $A_{Control}$, and the Test Well is marked as A_{Test} . Finally calculate $\Delta A_{Test} = A_{Test} - A_{Control}$, $\Delta A_{Standard} = A_{Standard} - A_{Blank}$.

Note: The Standard Well and Blank Well only need to be done once or twice, Each Test Well needs to be provided with a Control Well. In order to guarantee the accuracy of experimental results, need to do a pre-experiment with 2-3 samples. If ΔA_{Test} is less than 0.05, increase the sample quantity appropriately. If ΔA_{Test} is larger than $\Delta A_{Standard}$ of 2 µmol/mL, the supernatant can be further diluted with deionized water, the supernatant can be appropriately diluted with deionized water, the calculated result multiplied by the dilution factor, or decrease the sample quantity appropriately.

Data Analysis

Note: We provide you with calculation formulae, including the derivation process and final formula. The two are exactly equal. It is suggested that the concise calculation formula in bold is final formula.

1. Drawing of standard curve

With the concentration of the standard solution as the x-axis and the $\Delta A_{Standard}$ as the y-axis, draw the standard curve and obtain the standard equation. The determination of ΔA_{Test} is brought into the equation to get x (μ mol/mL).

2. Calculation of the S-ACX activity

Active unit definition: At 50°C and pH 4.8, the amount of enzyme needed to decompose xylan to produce 1 µmol of reducing sugar per gram of soil per hour was defined as one unit of enzyme activity.

S-ACX (U/g soli)=x×V_{Reaction}÷W÷T×F=0.075×x÷W×F

V_{Reaction}: Enzymatic reaction volume, 0.15 mL; T: reaction time, 2 h; W: weight of sample, g; F: dilution multiple.

Typical Data

The following data are for reference only. And the experimenters need to test the samples according to their own experiments.

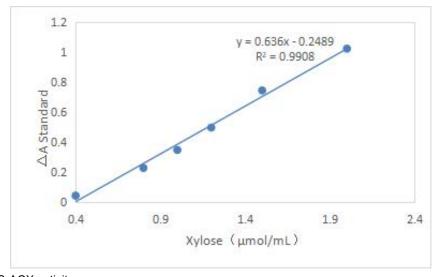


Figure 1. Standard curve of S-ACX activity.

Examples:

Take 0.02 g of fresh soil sample that has been dried in a 37° C oven and follow the steps described above, and after an 5-fold dilution, use 96-well plate to calculate ΔA_{Test} =0.797-0.076=0.721, x=1.524. The activity calculated according to the soil sample mass is as follows:

S-ACX (U/g soli)=0.075×1.524÷0.02×5=28.575 U/g.



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Recommended Products

Catalog No. Product Name	
KTB4012	CheKine™ Micro Soil Nitrate Nitrogen Assay Kit
KTB4014	CheKine™ Micro Acid Soil Available Phosphorous Assay Kit
KTB4041	CheKine™ Micro Soil Alkaline Phosphatase(S-AKP/ALP) Activity Assay Kit
KTB4050	CheKine™ Micro Soil Catalase (S-CAT) Activity Assay Kit

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes. For your safety and health, please wear a lab coat and disposable gloves.

